

Determination of Lead in TSP by Inductively Coupled Plasma Mass Spectrometry (ICP-MS) with Heated Ultrasonic Nitric and Hydrochloric Acid Filter Extraction

Standard Operating Procedure (SOP)

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EPA Contract No. EP-D-08-047

Office of Air Quality Planning and Standards
U.S. Environmental Protection Agency
Research Triangle Park, NC 27711

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The information in this document has been funded wholly or in part by the United States Environmental Protection Agency (USEPA) under EPA Contract No. EP-D-08-047. Mention of trade names or commercial products does not constitute endorsement or recommendation for use.

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1.0 SCOPE AND APPLICABILITY

Lead poisoning can cause irreversible brain damage and can impair mental functioning in children. It can retard mental and physical development and reduce attention span. In adults, it can cause irritability, poor muscle coordination, and nerve damage to the sense organs and nerves controlling the body.

Since 1980, lead emissions have decreased nearly 97 percent, according to EPA, and levels of lead in the air are currently much lower than they were in 1978. The drop in emissions is being attributed primarily to the phase-out of lead in gasoline. However, EPA estimates that over 1,300 tons of lead are emitted into the air annually from sources including smelters, iron and steel foundries, and general aviation gasoline.¹ Based on its review of the air quality criteria and national ambient air quality standards (NAAQS) for lead (Pb), EPA has made revisions to the primary and secondary NAAQS for Pb to protect public health and welfare. EPA has revised the level to $0.15 \mu\text{g}/\text{m}^3$ while it is retaining the current indicator of lead in total suspended particles (Pb-TSP).²

This Standard Operating Procedure (SOP) is for the Determination of Lead in TSP by Inductively Coupled Plasma Mass Spectrometry (ICP-MS) with Heated Ultrasonic Nitric and Hydrochloric Acid Filter Extraction.

NOTE: This SOP is based on USEPA's Office of Solid Waste (SW-846) Method 6020A – Inductively Coupled Plasma Mass Spectrometry.³ Wording in certain sections of this SOP are paraphrased or taken directly from Method 6020A.

- 1.1 Inductively coupled plasma mass spectrometry (ICP-MS) is applicable for the sub- $\mu\text{g}/\text{mL}$ determination of lead in a wide variety of matrices. This procedure describes a method for the acid extraction of lead in particulate matter collected on high-volume, glass fiber, TSP filters and measurement of the extracted lead using ICP-MS.
- 1.2 Due to variations in the isotopic abundance of lead, the value for total lead must be based on the sum of the signal intensities for isotopic masses, 206, 207, and 208. Most instrument software packages are able to sum the primary isotope signal intensities automatically.
- 1.3 ICP-MS requires the use of an internal standard. ^{115}In , ^{165}Ho , and ^{209}Bi are recommended internal standards for the determination of lead.
- 1.4 Use of this method is restricted to use by, or under supervision of, properly trained and experienced personnel. Requirements include training and experience

in inorganic sample preparation including acid extraction and also knowledge in the recognition and in the correction of spectral, chemical and physical interference in ICP-MS. Each analyst must demonstrate the ability to generate acceptable results with this method.

2.0 SUMMARY OF METHOD

- 2.1 This method describes the acid extraction of lead in particulate matter collected on 8 x 10 inch, glass fiber, ambient air filters using a high-volume TSP sampling device as described in 40 CFR Part 50, Appendix B⁵ with subsequent measurement of the dissolved lead by ICP-MS. The Method Detection Limit (MDL) (sensitivity) was demonstrated to 0.0000161 $\mu\text{g}/\text{m}^3$ using Pb-spiked filter strips analyzed in accordance with the guidance provided in 40 CFR Part 136, Appendix B⁶. The method range was demonstrated from 0.00120 $\mu\text{g}/\text{m}^3$ to 0.480 $\mu\text{g}/\text{m}^3$, based on the low and high calibration curve standards and a nominal filter volume of 2000 m^3 .
- 2.2 This SOP includes one extraction method. In this method, a solution of nitric and hydrochloric acids is added to the filter samples in plastic tubes and the tubes are placed in a heated ultrasonic bath for one hour to facilitate the extraction of lead. Following ultrasonication, the samples are brought to a final volume of 40mL, vortex mixed or shaken vigorously, and centrifuged prior to aliquots being taken for ICP-MS analysis.
- 2.3 Calibration standards and check standards are prepared to matrix match the acid composition of the samples. ICP-MS analysis is then performed. With this method, the samples are first aspirated and the aerosol thus created is transported by a flow of argon gas into the plasma torch. The ions produced (e.g., Pb^{+1}) in the plasma are extracted via a differentially-pumped vacuum interface and are separated on the basis of their mass-to-charge ratio. The ions are quantified by a channel electron multiplier or a Faraday detector and the signal collected is processed by the instrument's software. Interferences must be assessed and corrected for, if present.

3.0 DEFINITIONS

Pb – Elemental or ionic lead

ICP-MS - Inductively Coupled Plasma Mass Spectrometer

MDL – Method detection limit

RL – Reporting limit

RSD – Relative standard deviation

RPD – Relative percent difference
ICB – Initial calibration blank
CCB – Continuing calibration blank
ICV - Initial calibration verification
CCV – Continuing calibration verification
LLCV – Lower Level Calibration Verification, serves as LLICV and LLCCV
RB - Reagent blank
RBS -Reagent blank spike
MSDS – Material Safety Data Sheet
NIST – National Institute of Standards and Technology
D.I. water - Deionized water
RTI – RTI International
SOP – Standard Operating Procedure
SRM – NIST Standard Reference Material
CRM – Certified Reference Material
USEPA – U.S. Environmental Protection Agency
v/v – volume to volume ratio

4.0 INTERFERENCES

- 4.1 Reagents, glassware, plasticware, and other sample processing hardware may yield artifacts and/or interferences to sample analysis. If reagent blanks, filter blanks, or quality control blanks yield results above the detection limit the source of contamination must be identified. All containers and reagents used in the processing of the samples must be checked for contamination prior to sample extraction and analysis. Reagents shall be diluted to match the final concentration of the extracts and analyzed for lead. Labware shall be rinsed with 0.38M HNO₃/0.84M HCl extraction solution and the solution analyzed. Once a reagent or labware article, such as extraction tubes, from a manufacturer have been successfully screened, additional screening is not required unless contamination is suspected.
- 4.2 Isobaric elemental interferences in ICP-MS are caused by isotopes of different elements forming atomic ions with the same nominal mass-to-charge ratio (m/z) as the species of interest. There are no species found in ambient air that will result in isobaric interference with the three lead isotopes, 206, 207, and 208, being

measured. Polyatomic interferences occur when two or more elements combine to form an ion with the same mass-to-charge ratio as the isotope being measured. Lead is not subject to interference from common polyatomic ions and no correction is required.

- 4.3 The distribution of lead isotopes is not constant. The analysis of total lead should be based on the summation of signal intensities for the isotopic masses 206, 207, and 208. In most cases, the instrument software can perform the summation.
- 4.4 Physical interferences are associated with the sample nebulization and transport processes as well as with ion-transmission efficiencies. Dissolved solids can deposit on the nebulizer tip of a pneumatic nebulizer and on the interface skimmers of the ICP-MS. Nebulization and transport processes can be affected if a matrix component causes a change in surface tension or viscosity. Changes in matrix composition can cause significant signal suppression or enhancement. These interferences are compensated for through use of internal standards. Sample dilution will reduce the effects of high levels of dissolved salts, but calibration standards must be prepared in the extraction medium and diluted accordingly.

5.0 HEALTH AND SAFETY CAUTIONS

- 5.1 This method does not address all the possible safety issues associated with its use. The laboratory is responsible for maintaining a safe work environment and compliance with all OSHA regulations. Material Safety Data Sheets (MSDS) for all chemicals used should be readily available to all personnel involved with the procedure.
- 5.2 Concentrated nitric and hydrochloric acids are moderately toxic and extremely irritating to the skin. Use these reagents in a hood, and if eye and skin contact occurs, flush with large volumes of water. Always wear safety glasses or a shield for eye protection when working with these reagents. The component of this procedure requiring the greatest care is nitric acid (HNO_3). Nitric acid is a strong, corrosive, oxidizing agent that requires protection of the eyes, skin, and clothing. Items to be worn during use of this reagent include:
- Safety goggles (or safety glasses with side shields)
 - Acid resistant rubber gloves
 - A protective garment such as a laboratory apron. Nitric acid spilled on clothing will destroy the fabric and result in a hole; contact with the skin underneath will result in a burn.

It is also essential that an eye wash fountain or eye wash bottle be available during performance of this method. An eye wash bottle has a spout that covers the eye. If acid or any other corrosive gets into the eye, the water in this bottle is squirted onto the eye to wash out the harmful material. Eye washing should be performed with large amounts of water immediately after exposure. Medical help should be sought immediately after washing. If either acid, but especially nitric acid, is spilled onto the skin, wash immediately with large amounts of water. Medical attention is not required unless the burn appears to be significant. Even after washing and drying, the nitric acid may leave the skin slightly brown in color, this will heal and fade with time.

- 5.3 Lead (Pb) salts and lead solutions are toxic. Great care must be taken to ensure that samples and standards are handled properly; wash hands thoroughly after handling.
- 5.4 Care must be taken when using the ultrasonic bath as it is capable of causing mild burns. Chemists should refer to the safety guidance provided by the manufacturer of their specific equipment.

6.0 EQUIPMENT AND SUPPLIES

6.1 Apparatus

- 6.1.1 Thermo Scientific X-Series Inductively Coupled Plasma Mass Spectrometer (ICP-MS) or equivalent. The system must be capable of providing resolution better or equal to 1.0 amu at 10% peak height. The system must have a mass range from at least 7 to 240 AMU that allows for the application of the internal standard technique. For the measurement of lead, an instrument with a collision or reaction cell is not required.
- 6.1.2 Heated ultrasonic bath capable of maintaining a temperature of 80°C; VWR Model 750HT, 240W, or equivalent. Ultrasonic bath must meet the following performance criteria:
 - 1. Cut a strip of aluminum foil almost the width of the tank and double the depth.
 - 2. Lower the foil into the operating ultrasonic bath vertically until almost touching the bottom of the tank and hold for 10 seconds.
 - 3. Remove the foil from the tank and observe the distribution of perforations and small pin prick holes. The indentations should be fine and evenly distributed. The even distribution of indentations indicates the ultrasonic bath is acceptable for use.

6.1.3 Laboratory centrifuge, Beckman GS-6, or equivalent.

6.1.4 Vortex mixer, VWR Signature Digital Vortex Mixer, VWR Catalog No. 14005-824, or equivalent.

6.2 Materials and Supplies

- Argon gas supply, 99.99% purity or better. National Welders Microbulk, or equivalent.
- Plastic digestion tubes with threaded caps for extraction and storage, SCP Science *DigiTUBE*® Item # 010-500-063, or equivalent.
- Pipette, Rainin EDP2, 100 μ L, $\pm 1\%$ accuracy, $\leq 1\%$ RSD (precision), with disposable tips, or equivalent.
- Pipette, Rainin EDP2, 1000 μ L, $\pm 1\%$ accuracy, $\leq 1\%$ RSD (precision), with disposable tips, or equivalent.
- Pipette, Rainin EDP2, 1-10 mL, $\pm 1\%$ accuracy, $\leq 1\%$ RSD (precision), with disposable tips, or equivalent.
- Pipette, Thermo Lab Systems, 5 mL, $\pm 1\%$ accuracy, $\leq 1\%$ RSD (precision), with disposable tips, or equivalent.
- Plastic tweezers; VWR Catalog No. 89026-420, or equivalent.
- Laboratory marker.
- Ceramic knife, Kyocera LK-25, or equivalent.
- Blank labels or labeling tape, VWR Catalog No. 36425-045, or equivalent.
- Graduated cylinder, 1 L, VWR 89000-260, or equivalent.
- Volumetric flask, Class A, 1 L, VWR Catalog No. 89025-778, or equivalent.
- Millipore Element deionized water system, or equivalent, capable of generating Type I water (>17.9 M Ω -cm).
- High purity glass fiber filters for use as blank filters for determination of MDL and laboratory blanks, Whatman EPM2000, or equivalent.

7.0 REAGENTS AND STANDARDS

7.1 Reagent- or trace metals-grade chemicals must be used in all tests. Unless otherwise indicated, it is intended that all reagents conform to the specifications

of the Committee on Analytical Reagents of the American Chemical Society, where such specifications are available.

- 7.2 Concentrated nitric acid, 67-70%, SCP Science Catalog No. 250-037-177, or equivalent.
- 7.3 Concentrated hydrochloric acid, 33-36%, SCP Science Catalog No. 250-037-175, or equivalent.
- 7.4 Deionized water – All references to deionized water in the method refer to Type I deionized water with a resistivity >17.9 MΩ-cm.
- 7.5 Extraction solution (1.03M HNO₃ + 2.23M HCl). Prepare by adding 500mL of D.I. water to a 1000mL flask, adding 64.4mL of concentrated HNO₃ and 182mL of concentrated HCl, shaking to mix, allowing solution to cool, diluting to volume with reagent water, and inverting several times to mix. Extraction solution must be prepared at least weekly.
- 7.6 Standard stock solutions may be commercially purchased for each element or as a multi-element mix. Internal standards may be purchased as a mixed multi-element solution. The manufacturer's expiration date and storage conditions must be adhered to.
 - 7.6.1 Lead standard, 1000 µg/mL, NIST traceable, commercially available with certificate of analysis. High Purity Standards Catalog No. 100028-1, or equivalent.
 - 7.6.2 Indium standard, 1000 µg/mL, NIST traceable, commercially available with certificate of analysis. High Purity Standards Catalog No. 100024-1, or equivalent.
 - 7.6.3 Bismuth standard, 1000 µg/mL, NIST traceable, commercially available with certificate of analysis. High Purity Standards Catalog No. 100006-1, or equivalent.
 - 7.6.4 Holmium standard, 1000 µg/mL, NIST traceable, commercially available with certificate of analysis. High Purity Standards Catalog No. 100023-1, or equivalent.
 - 7.6.5 Second source lead standard, 1000 µg/mL, NIST traceable, commercially available with certificate of analysis. Must be from a different vendor or lot than 7.6.1. Inorganic Ventures Catalog No. CGPB-1, or equivalent.

7.6.6 Standard Reference Materials, NIST SRM 2583⁷, 2586⁸, 2587⁹ or 1648¹⁰, or equivalent.

Note: The In, Bi, and Ho internal standards may also be purchased as 10 µg/mL standards. Calibration standards are prepared by diluting stock standards to the appropriate levels in the same acid concentrations as in the final sample volume (0.38M HNO₃ + 0.84M HCl). The typical range for calibration standards is 0.005 to 2.00 µg/mL. At a minimum the curve must contain a blank and five (5) lead containing calibration standards. The calibration standards shall be stored at ambient laboratory temperature. Calibration standards must be prepared weekly and verified against a freshly prepared ICV using a NIST-traceable source different from the calibration standards.

7.7 Internal standards may be added to the test solution or by on-line addition. The nominal concentration for an internal standard is 0.010 µg/mL (10 ppb). Bismuth is the preferred internal standard for lead but holmium and indium may be used in the event the sample contains bismuth and high recoveries are observed.

7.8 Three laboratory blank solutions are required for analysis: (1) the calibration blank is used in the construction of the calibration curve and as a periodic check of system cleanliness (ICB and CCB); (2) the reagent blank (RB) is carried through the extraction process to assess possible contamination; and (3) the rinse blank is run between samples to clean the sample introduction system. If reagent blanks or laboratory blanks yield results above the detection limit, the source of contamination must be identified. Screening of labware and reagents is addressed in Section 4.1.

7.8.1 The calibration blank is prepared in the same acid matrix as the calibration standards (0.38M HNO₃ + 0.84M HCl) and samples and contains all internal standards used in the analysis.

7.8.2 The Reagent Blank (RB) contains all reagents used in the extraction and is carried through the extraction procedure at the same time as the samples.

7.8.3 The rinse blank is a solution of 1-2% nitric acid (v/v) in reagent grade water. A sufficient volume should be prepared to flush the system between all standards and samples analyzed.

7.8.4 EPA procures filters and distributes them to the state monitoring agencies collecting Pb in support of the National Ambient Air Quality Standard (NAAQS). If filter lot blanks are provided to the laboratory for analysis, consult 40CFR, Appendix G to Part 50, Section 6.1.1 for guidance on testing.

- 7.9 The Initial Calibration Verification (ICV), Lower Level Calibration Verification (LLCV), and Continuing Calibration Verification (CCV) solutions are prepared from a different lead source than the calibration curve standards and at a concentration that is either at or below the midpoint on the calibration curve, but within the calibration range. Both are prepared in the same acid matrix as the calibration standards. Note that the same solution may be used for both the ICV and CCV. The ICV/CCV and LLCV solutions must be prepared fresh daily.
- 7.10 Tuning Solution. Prepare a tuning solution according to the instrument manufacturer's recommendations. This solution will be used to verify the mass calibration and resolution of the instrument.

8.0 QUALITY CONTROL

- 8.1 Standard quality control practices shall be employed to assess the validity of the data generated. Included are: Method Detection Limit (MDL), Reporting Limit (RL), Reagent Blank (RB), duplicate samples, spiked samples, serial dilutions, ICV, CCV, LLCV, ICB, CCB, and SRMs/CRMs.
- 8.2 MDLs must be calculated by analyzing seven replicates of a known low-level spike added to a filter strip and carried through the entire extraction procedure. The test solutions shall be spiked at 2-5 times the estimated MDL. The MDL is defined as 3.143 times the standard deviation of the seven replicates in accordance with 40 CFR Part 136, appendix B⁶. The reporting limit, RL, is defined as the lowest calibration standard included in the curve or the LLCV concentration. Results should not be reported below this limit unless clearly marked as estimated. See Method 6020A, Section 10.4.3 for additional guidance.
- 8.3 For each batch of samples, one method or reagent blank (RB) and one method or reagent blank spike (RBS) spiked at the same level as the sample spike (see 8.6) must be prepared and carried throughout the entire process. The results of the RB must be below the RL. The recovery for the RBS must be within $\pm 20\%$ of the expected value. If the reagent blank yields a result above the RL, the source of contamination must be identified and the extraction and analysis repeated. Reagents and labware must be suspected as sources of contamination. Screening of reagents and labware is addressed in Section 4.1.
- 8.4 Any samples that exceed the highest calibration standard must be diluted and rerun so that the concentration falls within the curve. The minimum dilution will be 1 to 5 with a 0.38M HNO₃ and 0.84M HCl solution.
- 8.5 The internal standard response must be monitored during the analysis. If the internal standard response falls below 70% or rises above 120% of expected due

to possible matrix effects, the sample must be diluted and reanalyzed. The minimum dilution will be 1 to 5 with a 0.38M HNO₃ and 0.84M HCl solution. If the first dilution does not correct the problem, additional dilutions must be run until the internal standard falls within the specified range.

- 8.6 For every batch of samples prepared, there must be one duplicate and one spike sample prepared. The spike added is to be at a level that falls within the calibration curve, normally the midpoint of the curve. The initial plus duplicate sample must yield an RPD of ≤ 20 . The spike must be within $\pm 20\%$ of the expected value.
- 8.7 For each batch of samples, one extract must be diluted five fold and analyzed. The corrected dilution result must be within $\pm 10\%$ of the undiluted result. The sample chosen for the serial dilution shall have a concentration at or above 10X the lowest standard in the curve to ensure the diluted value falls within the curve. If the serial dilution fails, chemical or physical interference should be suspected.
- 8.8 ICB, ICV, LLCV, CCB and CCV samples are to be run as shown in the following table.

Sample	Frequency	Performance Specification
ICB	Prior to first sample	Below the RL
ICV	Prior to first sample	Within 90 to 110% of the expected value
LLCV	Daily, before first sample and after last sample	$\pm 30\%$ of the expected value
CCB	After every 10 extracted samples	Less than the RL
CCV	After every 10 extracted samples	Within 90-110% of the expected value

If any of these QC samples fails to meet specifications, the source of the unacceptable performance must be determined, the problem corrected, and any samples not bracketed by passing QC samples must be reanalyzed.

- 8.9 For each batch of samples, one certified reference material (CRM) must be combined with a blank filter strip and carried through the entire extraction procedure. A blank filter strip spiked with a NIST traceable solution different from that used for preparing the calibration standards or a solid CRM combined with a blank filter strip in the extraction tube are acceptable for use. The result must be within $\pm 20\%$ of the expected value.
- 8.10 For each run, a LLCV must be analyzed. The LLCV shall be prepared at a concentration not more than three times the lowest calibration standard and at a concentration not used in the calibration curve. The LLCV is used to assess performance at the low end of the curve. If the LLCV fails the run must be terminated, the problem corrected, the instrument recalibrated, and the analysis repeated.
- 8.11 Pipettes used for volumetric transfer must have the calibration checked at least once every 6 months and pass $\pm 1\%$ accuracy and $\leq 1\%$ RSD (precision) based on five replicate readings. The pipettes must be checked weekly for accuracy with a single replicate. Any pipette that does not meet $\pm 1\%$ accuracy on the weekly check must be removed from service, repaired, and pass a full calibration check before use.

9.0 CALIBRATION

Follow the instrument manufacturer's instructions for the routine maintenance, cleaning, and ignition procedures for the specific instrument being used.

- 9.1 Ignite the plasma and wait for at least one half hour for the instrument to warm up before beginning any pre-analysis steps.
- 9.2 For the Thermo X-Series with Xt cones, aspirate a 10 ng/mL tune solution containing In, Bi, and Ce. Monitor the intensities of In, Bi, Ce, and CeO and adjust the instrument settings to achieve the highest In and Bi counts while minimizing the CeO/Ce oxide ratio. For other instruments, follow the manufacturer's recommended practice. Tune to meet the instrument manufacturer's specifications. After tuning, place the sample aspiration probe into a 2% nitric acid rinse solution for at least 5 minutes to flush the system.
- 9.3 Aspirate a 5 ng/mL solution containing Co, In, and Bi to perform a daily instrument stability check. Run 10 replicates of the solution. The %RSD for the replicates must be less than 3% at all masses. If the %RSD is greater than 3%, the sample introduction system, pump tubing, and tune should be examined, and the analysis repeated. Place the sample aspiration probe into a 2% nitric acid rinse solution for at least 5 minutes to flush the system.

- 9.4 Load the calibration standards in the autosampler and analyze using the same method parameters that will be used to analyze samples. The curve must include one blank and at least 5 lead-containing calibration standards. The correlation coefficient must be at least 0.998 for the curve to be accepted. The lowest standard must recover $\pm 15\%$ of the expected value and the remaining standards must recover $\pm 10\%$ of the expected value to be accepted.
- 9.5 Immediately after the calibration curve is completed, analyze an ICV and an ICB. The ICV must be prepared from a different source of lead than the calibration standards. The ICV must recover 90-110 % of the expected value for the run to continue. The ICB must be less than the RL. If either the ICV or the ICB fails, the run must be terminated, the problem identified and corrected, and the analysis re-started.
- 9.6 A LLCV, CCV and a CCB must be run after the ICV and ICB. A CCV and CCB must be run at a frequency of not less than every 10 extracted samples. The CCV solution is prepared from a different source than the calibration standards and may be the same as the ICV solution. The LLCV must be within $\pm 30\%$ of expected value. The CCV value must be within $\pm 10\%$ of expected for the run to continue. The CCB must be less than the RL. If either the CCV, LLCV, or CCB fails, the run must be terminated, the problem identified and corrected, and the analysis re-started from the last passing CCV/LLCV/CCB set.
- 9.7 A LLCV, CCV, and CCB set must be run at the end of the analysis. The LLCV must be within $\pm 30\%$ of expected value. If either the CCV, LLCV, or CCB fails, the run must be terminated, the problem identified and corrected, and the analysis re-started from the last passing CCV/LLCV/CCB set.

10.0 FILTER STRIP EXTRACTION

All plasticware (e.g., Nalgene) and glassware used in the extraction procedures is soaked in 1% HNO₃ for at least 24 hours and rinsed with reagent water prior to use. All mechanical pipettes used must be calibrated to $\pm 1\%$ accuracy and $\leq 1\%$ RSD at a minimum of once every 6 months.

10.1 Sample Preparation – Heated Ultrasonic Bath

- 10.1.1 Using a ceramic knife and non-metal ruler, cut a 3/4 in. X 8 in. strip from the exposed area of the filter by cutting a strip from the edge of the filter where it has been folded along the 10” side at least 1 in. from the right or left side to avoid the un-sampled area covered by the filter holder. The filters must be carefully handled to avoid dislodging deposits.

10.1.2 Using plastic tweezers, roll the filter strip up in a coil like a cinnamon bun and place the rolled strip in the bottom of a labeled 50mL extraction tube (See Figure 1). In a fume hood, add 15.0 ± 0.15 mL of the extraction solution (Section 7.5) using a calibrated mechanical pipette. Ensure that the extraction solution completely covers the filter strip.

10.1.3 Loosely cap the 50mL extraction tube and place it upright in a plastic rack (See Figure 2). When all samples have been prepared, place the racks in an uncovered heated ultrasonic water bath that has been preheated to $80 \pm 5^\circ\text{C}$ and ensure that the water level in the ultrasonic is above the level of the extraction solution in the tubes but well below the level of the extraction tube caps to avoid contamination. Start the ultrasonic bath and allow the unit to run for 1 hour ± 5 minutes at $80 \pm 5^\circ\text{C}$.

10.1.4 Remove the rack(s) from the ultrasonic bath and allow the racks to cool.

10.1.5 Add 25.0 ± 0.25 mL of D.I. water with a calibrated mechanical pipette to bring the sample to a final volume of 40.0 ± 0.4 mL. Tightly cap the tubes and vortex mix or shake vigorously. Place the extraction tubes in an appropriate holder and centrifuge for 20 minutes at 2500RPM.

CAUTION- Make sure that the centrifuge holder has a flat bottom to support the flat bottomed extraction tubes.

10.1.6 Pour an aliquot of the solution into an autosampler vial for ICP-MS analysis to avoid the potential for contamination. Do not pipette an aliquot of solution into the autosampler vial.

10.1.7 Decant the extract to a clean tube, cap tightly, and store the sample extract at ambient laboratory temperature. Extracts may be stored for up to six months from the date of extraction.

11.0 MEASUREMENT PROCEDURE

11.1 Follow the instrument manufacturer's startup procedures for the ICP-MS.

11.2 Set instrument parameters to the appropriate operating conditions as presented in the instrument manufacturer's operating manual and allow the instrument to warm up for at least 30 minutes.

11.3 Calibrate the instrument per Section 9 of this SOP.

11.4 Verify the instrument is suitable for analysis as defined in Sections 9.2 and 9.3.

- 11.5 As directed in Section 8 of this SOP, analyze an ICV and ICB immediately after the calibration curve followed by a LLCV, then CCV and CCB. The acceptance requirements for these parameters are presented in Section 8.8.
- 11.6 Analyze a CCV and a CCB after every 10 extracted samples.
- 11.7 Analyze a LLCV, CCV and CCB at the end of the analysis.
- 11.8 Typical run samples will include field samples, field sample duplicates, spiked field sample extracts, serially diluted samples, the set of QC samples listed in 8.8 above, and one or more CRMs/SRMs.
- 11.9 Any samples that exceed the highest standard in the calibration curve must be diluted and reanalyzed so that the diluted concentration falls within the calibration curve.

12.0 RESULTS

- 12.1 The filter results must be reported in $\mu\text{g}/\text{mL}$ as analyzed. Any additional dilutions done must be accounted for in the instrument software. The internal standard recoveries must be included in the result calculation; this is done by the ICP-MS software for most all commercial instruments. Final results should be reported in $\mu\text{g Pb}/\text{m}^3$ to three significant figures as follows in 12.2.

12.2 $C = \frac{(\mu\text{g Pb}/\text{mL} \times 40\text{mL}/\text{strip} \times A)}{V_s}$

$$V_s$$

Where:

$$C = \text{Concentration, } \mu\text{g Pb} / \text{m}^3$$

$$\mu\text{g Pb}/\text{mL} = \text{Lead concentration in solution}$$

$$40.0 \text{ mL}/\text{strip} = \text{Total extraction solution volume}$$

$$A = \text{Area correction} = 63.0\text{in}^2 \text{ sampled} / 5.25\text{in}^2 \text{ analyzed} = 12.0$$

$$V_s = \text{Actual volume of air sampled}$$

The calculation assumes the use of a standard 8" X 10" TSP filter which has a sampled area of 9" X 7" (63.0in^2) due to the $\frac{1}{2}$ " filter holder border around the outer edge. The $\frac{3}{4}$ " X 8" strip has a sampled area of $\frac{3}{4}$ " X 7" (5.25in^2). If filter lot

blanks are provided for analysis, refer to 40CFR, Appendix G to Part 50, Section 6.1.1 for guidance on testing.

13.0 METHOD PERFORMANCE

Information in this section is an example of typical performance results achieved by following this SOP. Actual performance must be demonstrated by each individual laboratory and instrument.

- 13.1 Performance data has been collected to determine the method detection limit (MDL) for this method. MDLs were determined for the ultrasonic/ nitric and hydrochloric acid extraction method in accordance with 40 CFR Part 136, Appendix B⁶. Table 1 shows the MDLs determined from seven reagent/filter blank solutions and seven reagent/filter blank solutions spiked with low level lead at three times the estimated MDL. The MDLs are well below the USEPA requirement of 5% of the current Pb NAAQS or 0.0075 µg/m³.
- 13.2 Recovery tests with filter strips spiked with NIST SRMs were performed using the ultrasonic/ nitric and hydrochloric acid filter extraction method and measurement of the dissolved lead with ICP-MS. Table 2 shows recoveries with these standard reference materials. The recoveries for the SRMs are ≥ 90% at the 95% confidence level.

14.0 POLLUTION PREVENTION

- 14.1 Pollution prevention encompasses any technique that reduces or eliminates the quantity and/or toxicity of waste at the point of generation. Numerous opportunities for pollution prevention exist in laboratory operation. Whenever feasible, laboratory personnel should use pollution prevention techniques to address their waste generation. The sources of pollution generated with this procedure are waste acid extracts and lead-containing solutions.
- 14.2 For information about pollution prevention that may be applicable to laboratories and research institutions consult *Less is Better: Laboratory Chemical Management for Waste Reduction*, available from the American Chemical Society's Department of Government Relations and Science Policy, 1155 16th St. N.W., Washington, D.C., 20036, www.acs.org.

15.0 WASTE MANAGEMENT

- 15.1 Laboratory waste management practices must be conducted consistent with all applicable rules and regulations. Laboratories are urged to protect air, water, and land by minimizing all releases from hood and bench operations, complying with

the letter and spirit of any sewer and discharge permits and regulations, and by complying with all solid and hazardous waste regulation. For further information on waste management, consult *The Waste Management Manual for Laboratory Personnel* available from the American Chemical Society listed in Section 14.2.

- 15.2 Waste nitric acid, hydrochloric acid, and solutions containing these reagents and/or Pb must be placed in labeled bottles and delivered to a commercial firm that specializes in removal of hazardous waste.

16.0 REFERENCES

- ¹ (<http://www.thompson.com/public/index.jsp>) Last accessed 09/08/09.
- ² 40 CFR Parts 50, 51, 53, and 58, National Ambient Air Quality Standards for Lead; Final Rule.
- ³ Method 6020A – Inductively Coupled Plasma Mass Spectrometry. U.S. Environmental Protection Agency. Revision 1, February 2007
- ⁴ 40 CFR, Part 50, Appendix G - Reference Method for the Determination of Lead in Suspended Particulate Matter Collected From Ambient Air.
- ⁵ 40 CFR, Part 50, Appendix B — Reference Method for the Determination of Suspended Particulate Matter in the Atmosphere (High-Volume Method)
- ⁶ 40 CFR, Part 136, Appendix B — Definition and Procedure for the Determination of the Method Detection Limit – Revision 1.1
- ⁷ NIST, *Certificate of Analysis: Standard Reference Materials 2583, Trace Elements in Indoor Dust, Nominal 90 mg/kg Lead*, National Institute of Standards and Technology, Gaithersburg, MD, 1998.
- ⁸ NIST, *Certificate of Analysis: Standard Reference Materials 2586, Trace Elements in Soil. Nominal 500 mg/Kg Lead*, National Institute of Standards and Technology, Gaithersburg, MD, 2008.
- ⁹ NIST, *Certificate of Analysis: Standard Reference Materials 2587, Trace Elements in Soil Containing Lead from Paint, Nominal 3000 mg/Kg Lead*, National Institute of Standards and Technology, Gaithersburg, MD, 2008.
- ¹⁰ NIST, *Certificate of Analysis: Standard Reference Materials 1648, Urban Particulate Matter, 0.655±0.033% Lead*, National Institute of Standards and Technology, Gaithersburg, MD, 2008.

17.0 FIGURES AND TABLES



Figure 1. Top view of filter strip rolled and placed in 28mm diameter plastic extraction tube.

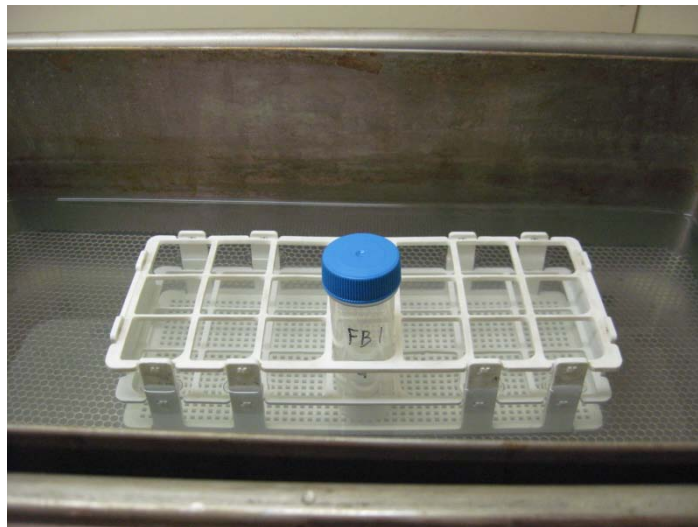


Figure 2. Filter in plastic extraction tube in tube rack placed in ultrasonic bath.

Table 1 . Method Detection Limits Determined by Analysis of Reagent/Glass Fiber Filter Blanks and Reagent/Glass Fiber Filter Blanks Spiked with Low-level Pb Solution.

Ultrasonic Extraction Method	Blank ($\mu\text{g}/\text{m}^3$)*	Pb-spiked ($\mu\text{g}/\text{m}^3$)*
n=1	0.0000434	0.0000702
n=2	0.0000420	0.0000715
n=3	0.0000439	0.0000611
n=4	0.0000407	0.0000587
n=5	0.0000437	0.0000608
n=6	0.0000437	0.0000607
n=7	0.0000403	0.0000616
Average	0.0000425	0.0000635
Standard Deviation	0.0000015	0.0000051
MDL**	0.0000047	0.0000161

* Assumes 2000 m³ air sample.

** MDL is 3.143 times the standard deviation of the results for seven consecutive sample replicates analyzed.

Table 2. Recoveries of Lead from NIST SRMs Spiked onto TSP Glass Fiber Filters.

Extraction Method	Recovery, ICP-MS, (%)		
	NIST 2586 Soil	NIST 2587 Soil	NIST 1648 Dust
Ultrasonic Bath	89.8±1.7	96.8±5.3	93.2±3.6