

Tuberculosis Testing and Laboratory Updates

Arizona State Public Health Laboratory

Stephanie Kreis
Arizona Department of Health Services
Bureau of State Laboratory Services
Bacteriology/Mycobacteriology/Parasitology
kreiss@azdhs.gov

Leadership for a Healthy Arizona



Overview

- Description of laboratory services at Arizona State Laboratory (ASL)
- Specimen collection and transport
- Testing workflow
- Tests available

Arizona State Laboratory

- Diagnostic and Reference laboratory
- Hours of Operation:
Monday – Friday, 8:00am
-5:00pm
- AZDHS Website
[http://www.azdhs.gov/
lab/micro/index.htm](http://www.azdhs.gov/lab/micro/index.htm)
 - Guide to Laboratory Services
 - Laboratory Submission form

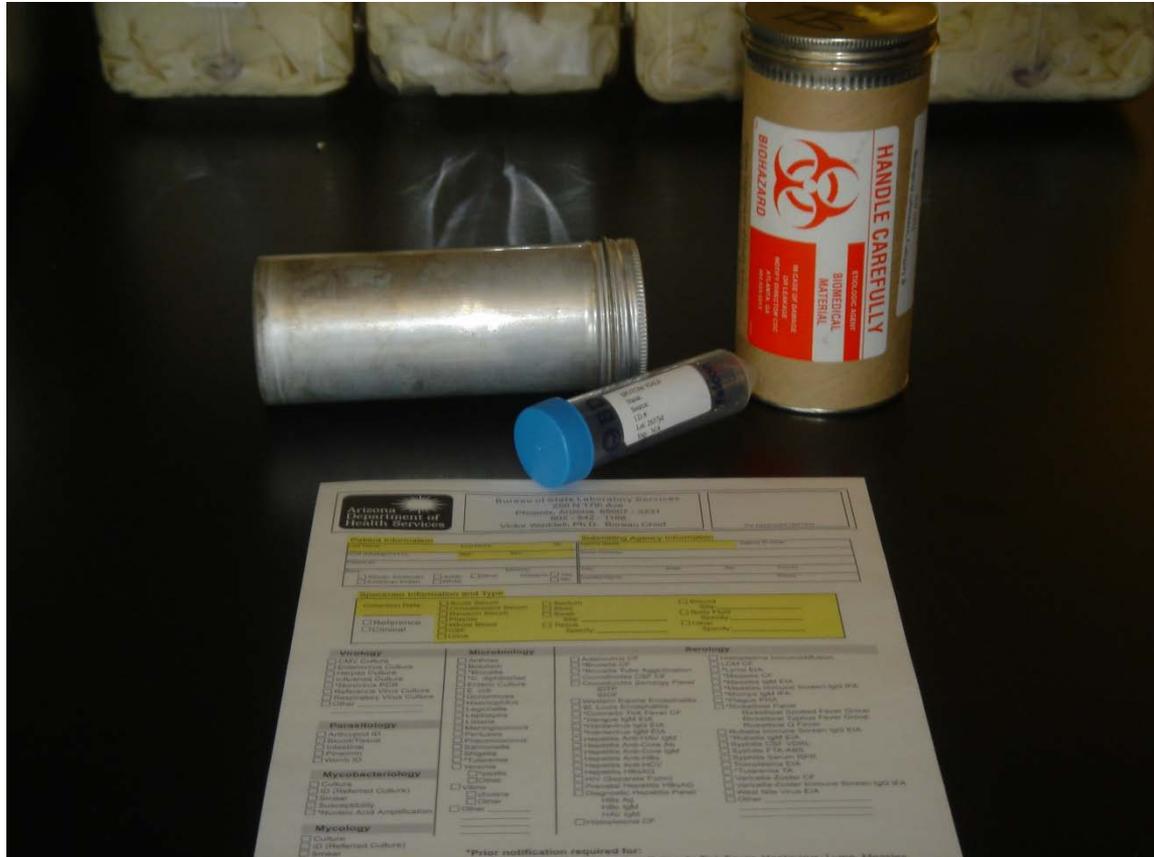


Arizona State Laboratory TB Services

- Processing of Diagnostic Specimens for AFB smear and culture
- AFB Microscopy
- AFB Culture and Identification
- Rapid testing: i.e. NAAT
- TB Drug Susceptibility testing
 - Broth method (rapid)
 - Agar Proportion method

Specimen Collection

- Specimen collection kits and mailing containers are provided by ASL
 - Contact by mail, phone, or fax
 - Arizona Department of Health Services
Bureau of State Laboratory Services
ATTN: Receiving Section
250 N. 17th Ave
Phoenix, AZ 85007
Fax: (602) 542-0760
Phone: (602) 542-1190



Example of Collection Kit Supplied by ASL
Inner metal screw capped container, outer screw capped cardboard container, place submission form around the outside of the inner metal container. **Label the specimen collection container with patient name and collection date.**

Specimen Collection

- Specimen collection containers requirements
 - Sterile
 - Leak-proof
 - Disposable
 - Non-breakable
 - Appropriately labeled!!!
- Laboratory Submission Form
 - <http://www.azdhs.gov/lab/micro/pdf/micro-submission-form.pdf>

Specimen Collection

- Quality specimens are vital for lab diagnosis of TB
- Collect aseptically, or bypass contamination as much as possible
- Avoid contamination with tap water (NTM may be present)
- Collect prior to therapy if possible
- No swabs, fixatives, preservatives

Laboratory Submission Form



Bureau of Laboratory Services

250 N. 17th Avenue Phoenix, Arizona 85007-3231

Tel: (602) 542-1188 Fax: (602) 364-0758

Victor Waddell, Ph.D., Bureau Chief

For Department Use Only

PATIENT INFORMATION

Last name: _____ First name: _____ MI: _____
DOB (MM/DD/YYYY): _____ Age: _____ Sex: M F T Patient ID: _____
Street address: _____ City: _____ State: _____ Zip: _____ County: select one
Ethnicity: Hispanic Yes No Race: White African American Asian American Indian/Alaska Native Other

SUBMITTING AGENCY INFORMATION

Agency name: _____ Agency ID Code: _____
Street address: _____ City: _____ State: _____ Zip: _____ County: select one
Contact name: _____ Tel: _____

ORDERING PROVIDER INFORMATION

Provider name: _____ Tel: _____
Facility name: _____ Tel: _____
Street address: _____ City: _____ State: _____ Zip: _____ County: select one

SPECIMEN INFORMATION & TYPE

Collection date: _____

<input type="checkbox"/> Serum <input type="checkbox"/> Acute <input type="checkbox"/> Convalescent	<input type="checkbox"/> Plasma	<input type="checkbox"/> Whole blood	<input type="checkbox"/> CSF
<input type="checkbox"/> Swab, site: _____	<input type="checkbox"/> Urine	<input type="checkbox"/> Sputum	<input type="checkbox"/> Stool
Other, specify: _____	<input type="checkbox"/> Tissue, specify: _____	<input type="checkbox"/> Wound, site: _____	<input type="checkbox"/> Body fluid, specify: _____

Clinical Reference: Broth Isolate Reason for testing: Outbreak Surveillance Post Mortem Screening Diagnostics

Outbreak name: _____

Laboratory Submission Form

LABORATORY NAME: _____

<p>VIROLOGY</p> <p><input type="checkbox"/> CMV Culture</p> <p><input type="checkbox"/> Enterovirus Culture</p> <p><input type="checkbox"/> Herpes Culture</p> <p><input type="checkbox"/> Influenza</p> <p><input type="checkbox"/> Norovirus PCR</p> <p><input type="checkbox"/> Reference Virus Culture</p> <p><input type="checkbox"/> Respiratory Virus Culture</p> <p><input type="checkbox"/> Other: _____</p> <hr/> <p>SELECT AGENTS**</p> <p><input type="checkbox"/> Avian Influenza H5N1</p> <p><input type="checkbox"/> *Bacillus anthracis</p> <p><input type="checkbox"/> *Brucella spp.</p> <p><input type="checkbox"/> *Burkholderia spp.</p> <p><input type="checkbox"/> *Francisella tularensis</p> <p><input type="checkbox"/> *Orthopox</p> <p><input type="checkbox"/> *Q Fever</p> <p><input type="checkbox"/> *SARS</p> <p><input type="checkbox"/> *Yersinia pestis</p> <p><small>**Please refer to the Guide to Laboratory Services: Microbiology, Section 8 for the definition of select agents and the testing available at Arizona State Laboratory</small></p>	<p>BACTERIOLOGY</p> <p><input type="checkbox"/> *Bordetella pertussis</p> <p><input type="checkbox"/> Campylobacter spp.</p> <p><input type="checkbox"/> *Clostridium botulinum toxin</p> <p><input type="checkbox"/> *Corynebacterium diphtheriae</p> <p><input type="checkbox"/> Enteric culture</p> <p><input type="checkbox"/> Escherichia coli / Shigatoxin</p> <p><input type="checkbox"/> Haemophilus influenzae</p> <p><input type="checkbox"/> Legionella spp.</p> <p><input type="checkbox"/> Leptospira spp.</p> <p><input type="checkbox"/> Listeria spp.</p> <p><input type="checkbox"/> Neisseria gonorrhoeae</p> <p><input type="checkbox"/> Neisseria meningitidis</p> <p><input type="checkbox"/> Salmonella spp.</p> <p><input type="checkbox"/> Shigella spp.</p> <p><input type="checkbox"/> Vibrio</p> <p><input type="checkbox"/> VISA/VRSA</p> <p><input type="checkbox"/> Yersinia (Non-pestis) Culture</p> <p><input type="checkbox"/> Other: _____</p> <hr/> <table border="0"> <tr> <td style="vertical-align: top;"> <p>PARASITOLOGY†</p> <p><input type="checkbox"/> Arthropod ID</p> <p><input type="checkbox"/> Blood/Tissue</p> <p><input type="checkbox"/> Intestinal</p> <p><input type="checkbox"/> Pinworm</p> <p><input type="checkbox"/> Worm ID</p> <p>† For malaria testing please collect patient travel history</p> </td> <td style="vertical-align: top;"> <p>MYCOBACTERIOLOGY</p> <p><input type="checkbox"/> Culture</p> <p><input type="checkbox"/> ID (Referred Culture)</p> <p><input type="checkbox"/> *Nucleic Acid Amplification</p> <p><input type="checkbox"/> Smear</p> <p><input type="checkbox"/> Susceptibility</p> </td> </tr> </table>	<p>PARASITOLOGY†</p> <p><input type="checkbox"/> Arthropod ID</p> <p><input type="checkbox"/> Blood/Tissue</p> <p><input type="checkbox"/> Intestinal</p> <p><input type="checkbox"/> Pinworm</p> <p><input type="checkbox"/> Worm ID</p> <p>† For malaria testing please collect patient travel history</p>	<p>MYCOBACTERIOLOGY</p> <p><input type="checkbox"/> Culture</p> <p><input type="checkbox"/> ID (Referred Culture)</p> <p><input type="checkbox"/> *Nucleic Acid Amplification</p> <p><input type="checkbox"/> Smear</p> <p><input type="checkbox"/> Susceptibility</p>	<p>SEROLOGY</p> <p><input type="checkbox"/> *Borrelia burgdorferi EIA (Lyme)</p> <p><input type="checkbox"/> *Brucella Tube Ag.</p> <p><input type="checkbox"/> Coccidioides Serology Panel <input type="checkbox"/> IDTP <input type="checkbox"/> IDCF</p> <p><input type="checkbox"/> Dengue IgG EIA</p> <p><input type="checkbox"/> Diagnostic Hepatitis Panel EIA <input type="checkbox"/> HBsAG <input type="checkbox"/> HBcIgM <input type="checkbox"/> HAV IgM</p> <p><input type="checkbox"/> *Francisella tularensis Tube Ag.</p> <p><input type="checkbox"/> *Hantavirus IgG EIA</p> <p><input type="checkbox"/> *Hantavirus IgM EIA</p> <p><input type="checkbox"/> Hepatitis Anti-HAV IgM</p> <p><input type="checkbox"/> Hepatitis Anti-Core IgM</p> <p><input type="checkbox"/> Hepatitis Anti-HCV</p> <p><input type="checkbox"/> Hepatitis HBsAG</p> <p><input type="checkbox"/> *Measles IgM EIA</p> <p><input type="checkbox"/> Mumps IgM EIA</p> <p><input type="checkbox"/> Rickettsial Panel IFA <input type="checkbox"/> Rickettsial Q Fever <input type="checkbox"/> Rickettsial Spotted Fever <input type="checkbox"/> Rickettsial Typhus Fever</p> <p><input type="checkbox"/> *Rubella IgM EIA</p> <p><input type="checkbox"/> St. Louis Encephalitis EIA</p> <p><input type="checkbox"/> Treponema pallidum CSF VDRL</p> <p><input type="checkbox"/> Treponema pallidum Serum TP-PA, RPR</p> <p><input type="checkbox"/> Western Equine Encephalitis EIA</p> <p><input type="checkbox"/> West Nile Virus EIA</p> <p><input type="checkbox"/> *Yersinia pestis PHA</p> <p><input type="checkbox"/> Other: _____</p>
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Submitting Lab Findings or Preliminary ID: _____

*Prior notification is required for: Bacillus anthracis, Bordetella pertussis, Brucella spp., Clostridium botulinum toxin, Corynebacterium diphtheriae, emerging or exotic diseases, Francisella tularensis, Hantavirus, Borrelia burgdorferi (Lyme), Measles, Mycobacteria NAA, Rubella, or Yersinia pestis testing. CALL: (602) 364-3676

ALL FIELDS HIGHLIGHTED IN YELLOW ARE REQUIRED FOR SPECIMEN PROCESSING. IN ADDITION, AT LEAST ONE TEST MUST BE REQUESTED.

Patient address and telephone number are required, when available, per R9-6-204(B3) <http://www.azdhs.gov/lab/micro/index.htm>

Updated 10/20/2010



Laboratory Submission Form

- Complete all highlighted (required) fields

PATIENT INFORMATION

Last name: _____ First name: _____ MI: _____
 DOB (MM/DD/YYYY): _____ Age: _____ Sex: M F T Patient ID: _____
 Street address: _____ City: _____ State: _____ Zip: _____ County: select one
 Ethnicity: Hispanic Yes No Race: White African American Asian American Indian/Alaska Native Other

SUBMITTING AGENCY INFORMATION

Agency name: _____ Agency ID Code: _____
 Street address: _____ City: _____ State: _____ Zip: _____ County: select one
 Contact name: _____ Tel: _____

ORDERING PROVIDER INFORMATION

Provider name: _____ Tel: _____
 Facility name: _____
 Street address: _____ City: _____ State: _____ Zip: _____ County: select one

SPECIMEN INFORMATION & TYPE

Collection date: _____

<input type="checkbox"/> Serum <input type="checkbox"/> Acute <input type="checkbox"/> Convalescent	<input type="checkbox"/> Plasma	<input type="checkbox"/> Whole blood	<input type="checkbox"/> CSF
<input type="checkbox"/> Swab, site: _____	<input type="checkbox"/> Urine	<input type="checkbox"/> Sputum	<input type="checkbox"/> Stool
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Clinical Reference: Broth Isolate Reason for testing: Outbreak Surveillance Post Mortem Screening Diagnostics
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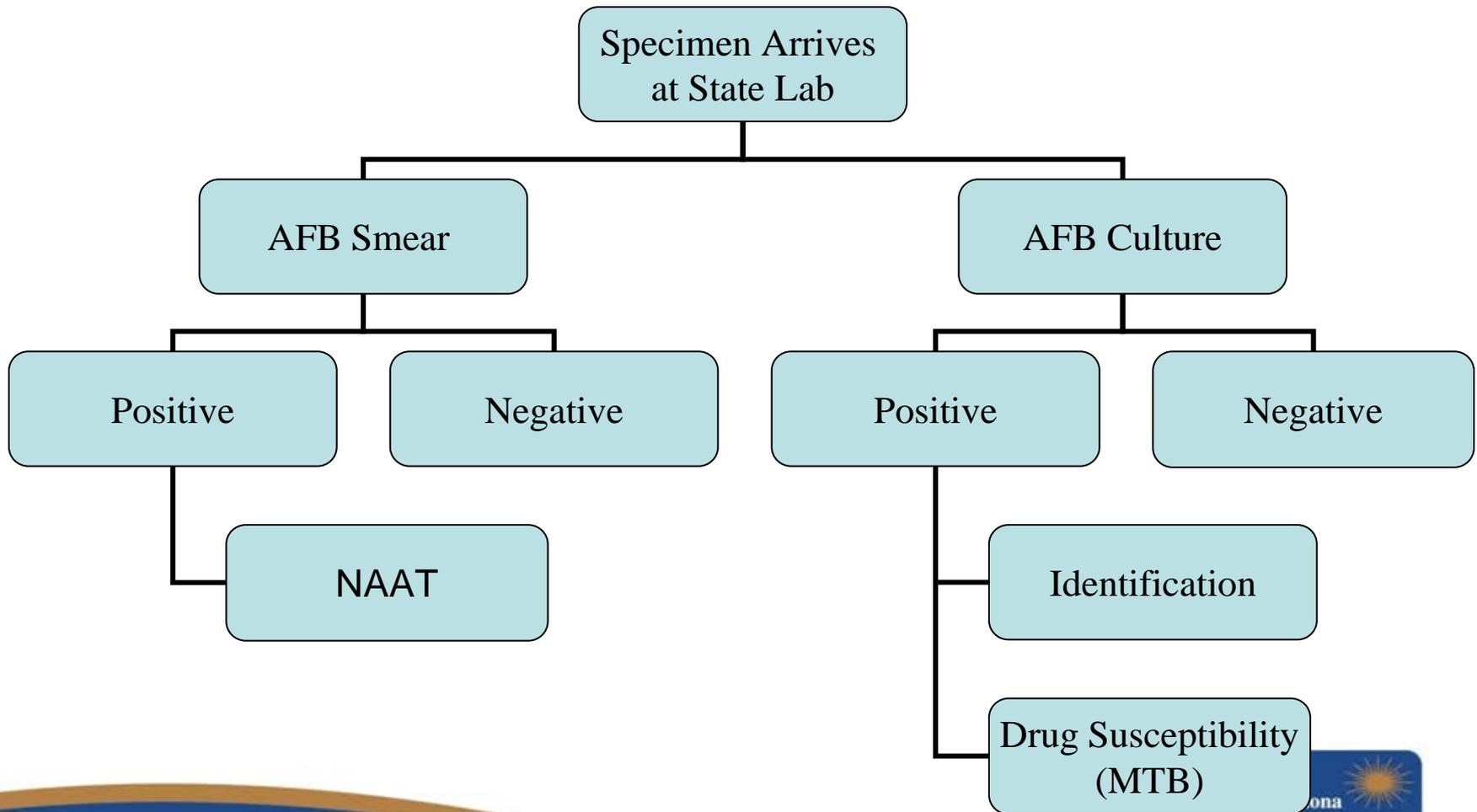
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AFB Specimen Transport

- Transport in as short as a time as possible to avoid overgrowth of contaminating bacteria
- Specimens that cannot be transported to the lab immediately should be refrigerated
- Reference the Guide to Laboratory Services
 - <http://www.azdhs.gov/lab/micro/index.htm>
- CDC recommendation:
 - Specimen received at laboratory within 24 hours of specimen collection.

Typical Specimen Workflow at ASL



AFB Smear Microscopy

- Quick and inexpensive
- Results should be available within 24 hours of receipt of specimen.
- Low sensitivity, 50-70% for pulmonary TB
- Not specific for MTBC
- Does not distinguish between live and dead bacilli

AFB Culture

- More sensitive than AFB smear
- Current recommendations are to use at least two types of media to maximize the recovery of mycobacterium
- Cultures monitored 6-8 weeks.



AFB Culture

- Liquid media typically provides more rapid results
- Combination of solid and liquid media maximizes recovery and provides an opportunity to look at colony morphology



Nucleic Acid Amplification Tests for Direct Detection of MTBC

- FDA-approved for use with respiratory specimens
- Non-FDA approved tests (RUO or not available in U.S.)
- Laboratory developed tests or LDT (e.g., DNA sequencing, Loop-mediated isothermal amplification [LAMP], and real-time PCR assays including molecular beacons)

Nucleic Acid Amplification Tests

- NAAT can detect MTBC genetic material directly from specimen within hours.
- Does not distinguish live and dead bacilli
- Results:
 - *Mycobacterium tuberculosis* complex detected
 - *Mycobacterium tuberculosis* complex not detected
 - Negative result does not necessarily mean absence of MTBC
 - Inhibition of amplification
 - Target below limit of detection

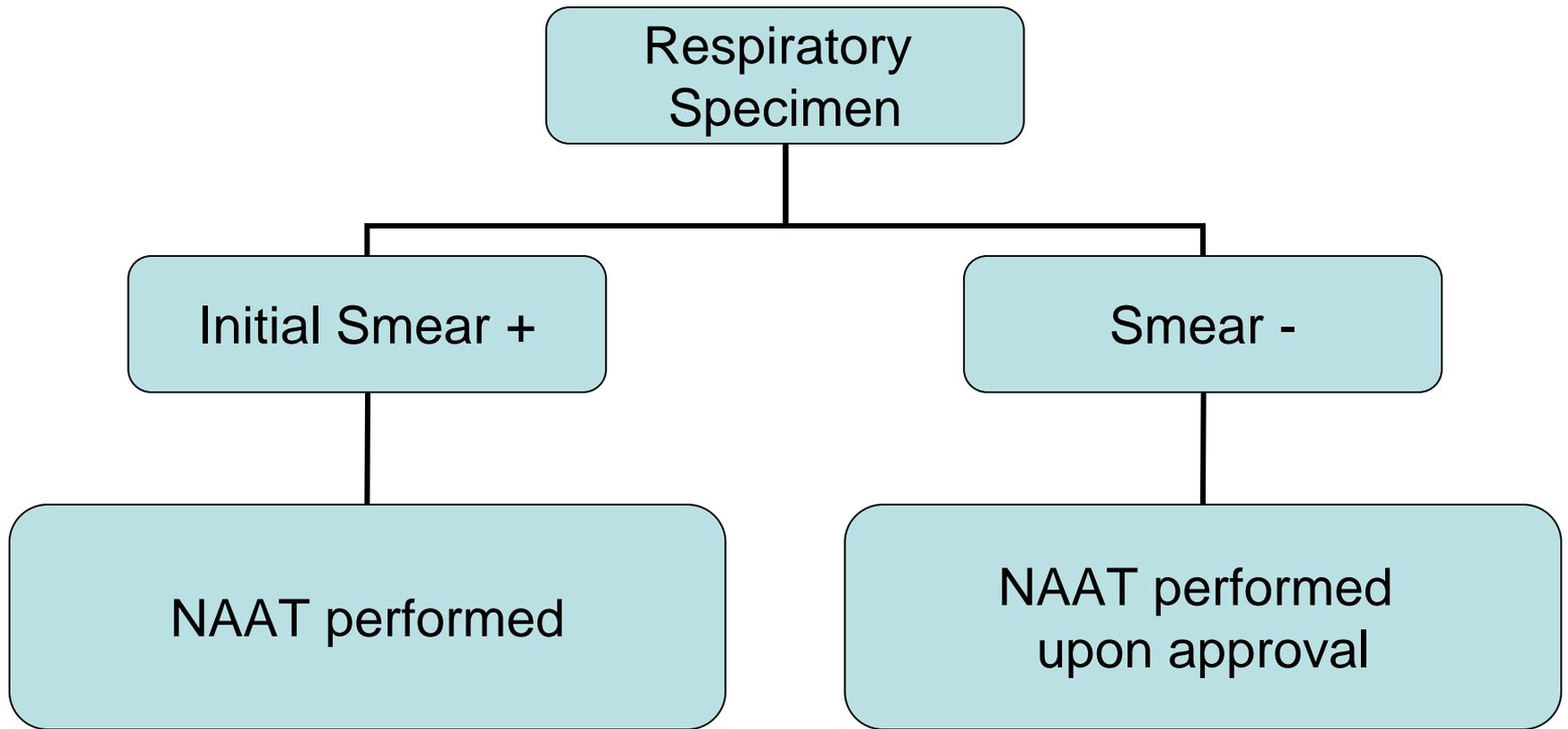
Nucleic Acid Amplification Tests

- Recommended Turnaround time: 24-48 hours from specimen receipt
- Most can be performed from smear positive or smear negative specimens
 - Sensitivity decreases for smear negative specimens
- Does not replace the need for culture confirmation. Culture still needed for conventional drug susceptibility testing and genotyping.

Updated Guidelines for the Use of NAAT in the Diagnosis of TB

- “NAAT should be performed on at least one respiratory specimen from each patient with signs and symptoms of pulmonary TB for whom a diagnosis of TB is being considered but has not yet been established, and for whom the test result would alter case management or TB control activities.” MMWR, 2009, 58:7-10
- State TB control program and ASL collaborated to determine criteria for testing.

ASL's NAAT Algorithm



How to get approval for NAAT?

- Contact State TB control program
- Approval will be based on whether the patient meets the following criteria
 - Have a cavitory lesion seen on CXR
 - Are in a long-term care facility
 - Are HIV positive
 - Are immunocompromised
 - Are on dialysis, or
 - If it will make a difference in the treatment/isolation of the patient

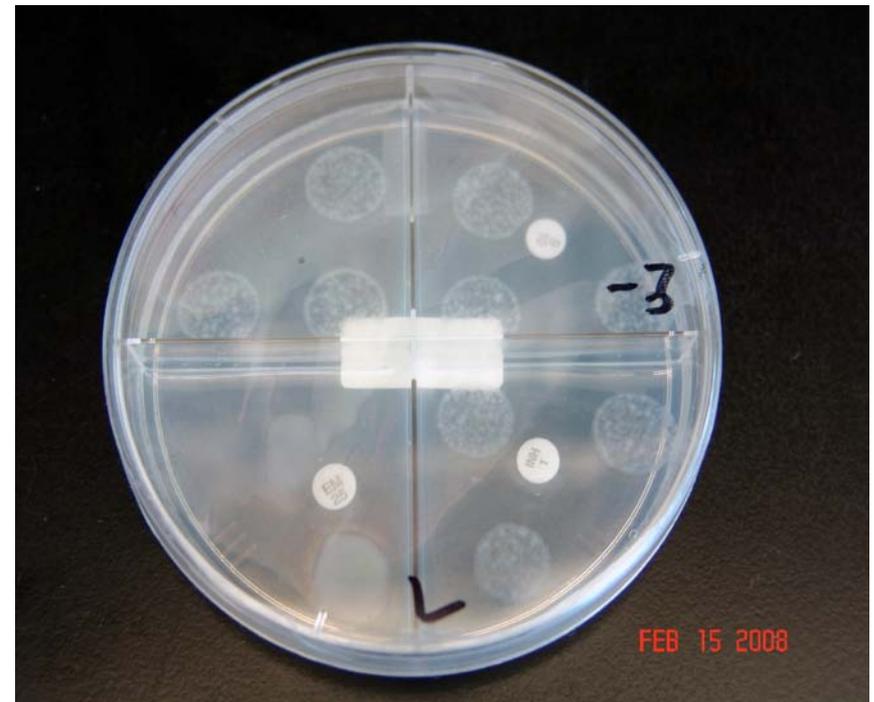
TB Drug Susceptibility Testing

- Broth Systems
 - Rapid results
 - Testing done from culture
 - FDA approved for first line TB drugs
 - Isoniazid, Rifampin, Ethambutol, PZA
 - Streptomycin
 - Recommended on all initial cases of MTBC
 - Should be repeated if there is clinical evidence of failure to respond to treatment or if cultures fail to convert to negative after 3 months of treatment.

Proportion Agar Drug Susceptibility Testing

- Proportion Agar Method
 - “Gold Standard” method used in the United States for several decades.
 - Equal quantities of several dilutions of a standard inoculum onto a agar based medium with and without drug
 - Number of CFU growing on drug containing quadrants are compared to those on the drug free medium and expressed as a percentage
 - Turnaround time: 21-28 days
- If drug resistance is suspected contact ASL or State TB control program.

Proportion Agar Drug Susceptibility Testing



Molecular Detection of Drug Resistance

- Laboratory developed tests (LDT)
- Non-FDA approved tests (Research Use Only [RUO] or not available in U.S.)

Molecular Detection of Drug Resistance

- The CDC began offering Molecular Detection of Drug Resistance for *Mycobacterium tuberculosis*, Sept 2009.
- PCR-based DNA sequencing for drug resistance
- Criteria for accepting specimens
 - High risk patients (RIF-R, MDR-TB)
 - High profile patients
 - Known RIF-R
 - Mixed or non-viable cultures

How to request MDDR services?

- Contact State TB control program to make a request
- ASL will contact the CDC for approval to send the specimen
- ASL will arrange to ship the specimen to the CDC
- Preliminary report issued with molecular results
- Final report issued upon completion of agar proportion drug susceptibility testing

CDC Molecular Detection of Drug Resistance Service

- Drugs and Genes offered with MDDR service (CDC)

Drug

Gene

RIF

rpoB

INH

inhA, katG

FQ

gyrA

KAN

rrs, eis

AMK

rrs

CAP

rrs, tlyA

EMB

embB

PZA

pncA

National TB Genotyping Program

- Began in January, 2004
- Goal: Genotype at least one isolate for every culture-positive case of TB
- Arizona State Lab actively participates in program by routinely submitting isolates to the assigned genotyping laboratory.

National TB Genotyping Program

- Results are uploaded to a national database.
- Results are used to
 - Confirm suspected links
 - Detect unsuspected transmission
 - Detect false positive cultures

Questions???



Leadership for a Healthy Arizona

